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Understanding how the physico-chemical properties of lipid-based drug carriers shape their interactions in complex biological environments

Anna Salvati

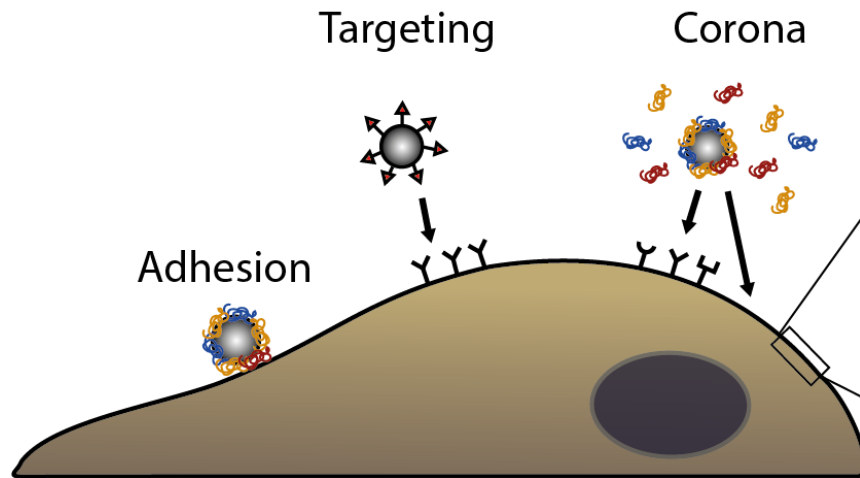
**Department of Nanomedicine & Drug Targeting
Groningen Research Institute of Pharmacy
University of Groningen**

**<http://www.rug.nl/staff/a.salvati/>
a.salvati@rug.nl**

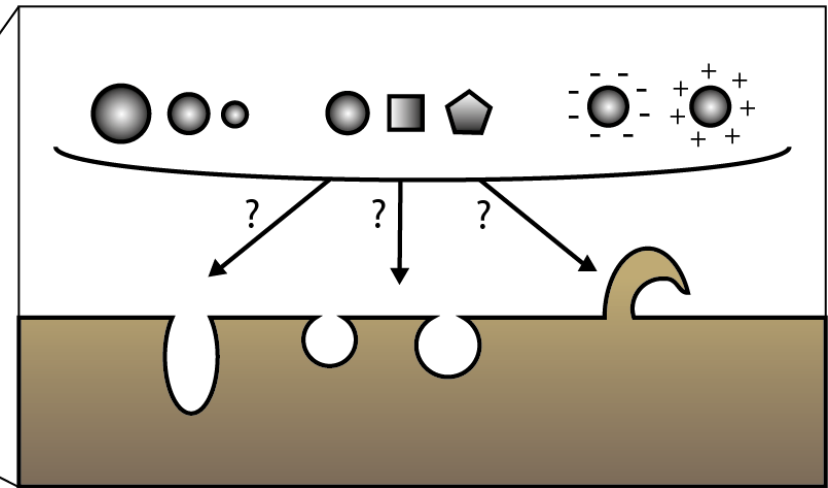
Nanomedicine for drug delivery

Trying to understand how nanomedicines interact with cells and how to control it
→ *“the cell biology of nanomedicine”*

1) Interactions at the cell membrane (receptors and targeting)



2) Uptake and intracellular processing



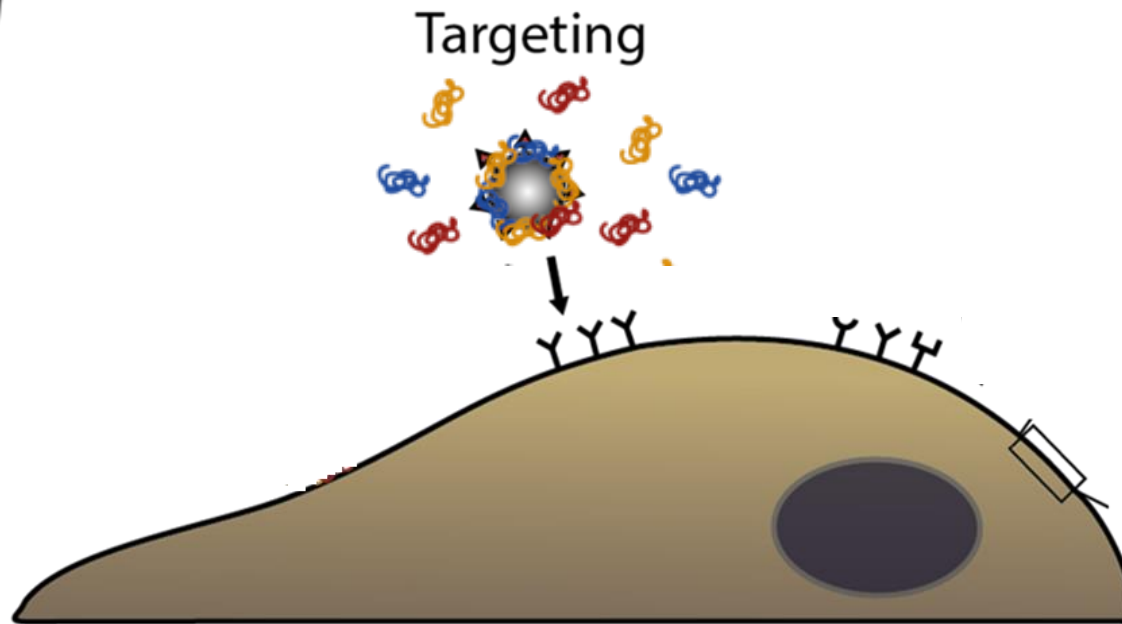
We need to learn the language to make it better!

How do nanomedicine physico-chemical properties shape their biological behavior?

Can we control the interaction with cell receptors?



- Changing nanomedicine physico-chemical properties
- Active targeting ligands on nanomedicines to interact with specific cells



**but upon administration: interactions with biological fluids and corona formation
→ BEFORE interactions with cells!**

***What are the implications for the interactions with cells?
...some examples***

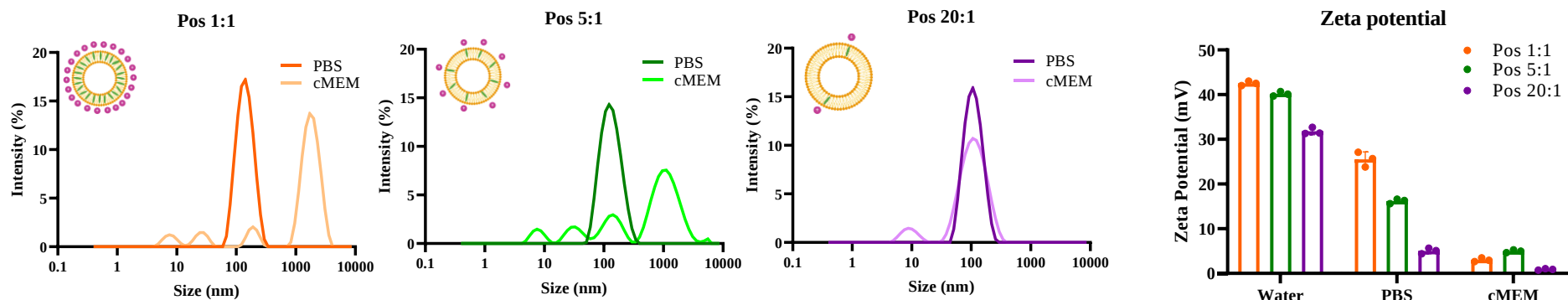
Why do positive nanoparticles usually have higher uptake than negative ones?

Feng Zhao

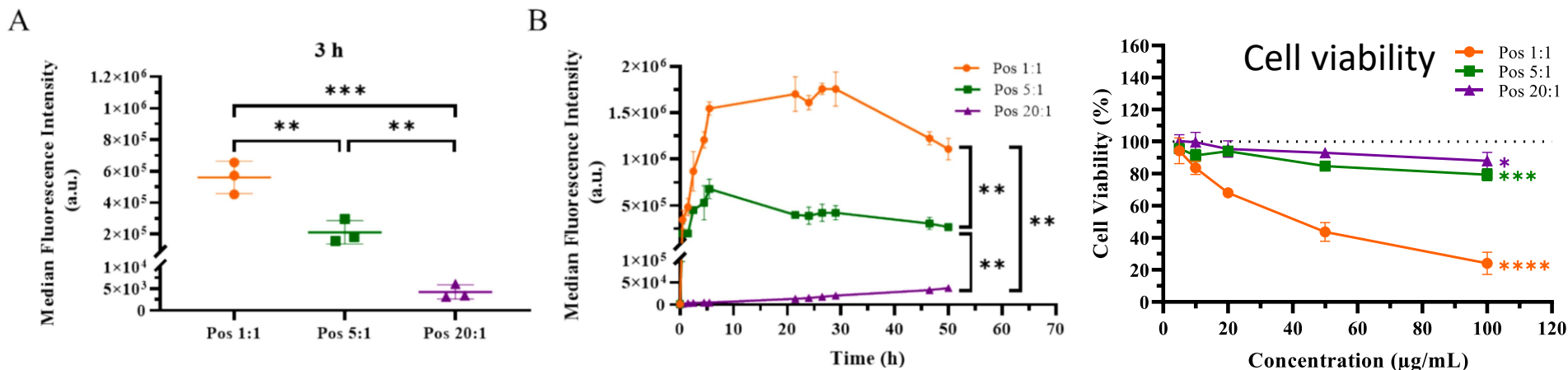


Optimization of DOPC-Dc-Chol liposome formulation

→ to find a non-toxic and stable positive liposome



Pos 1:1 and 5:1 aggregate in the medium with 10% FBS, 20:1 is stable

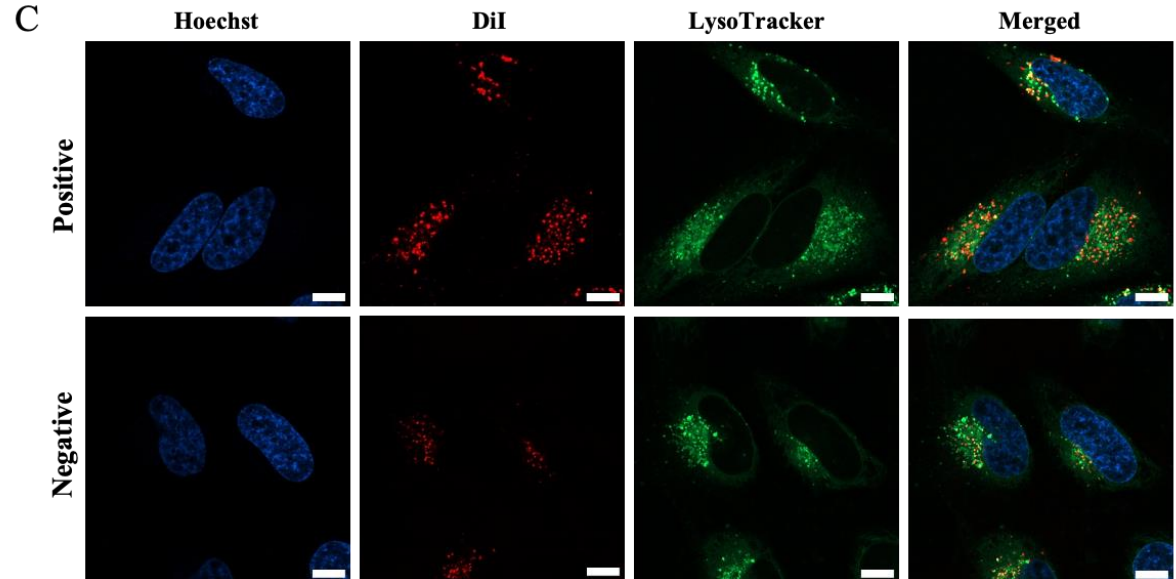
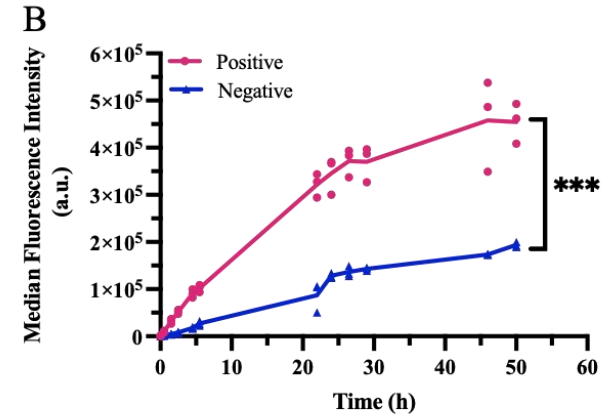
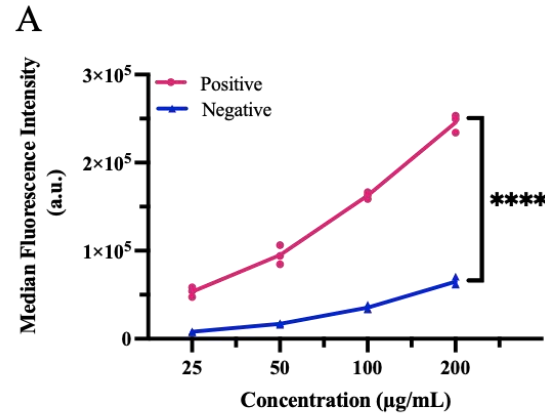
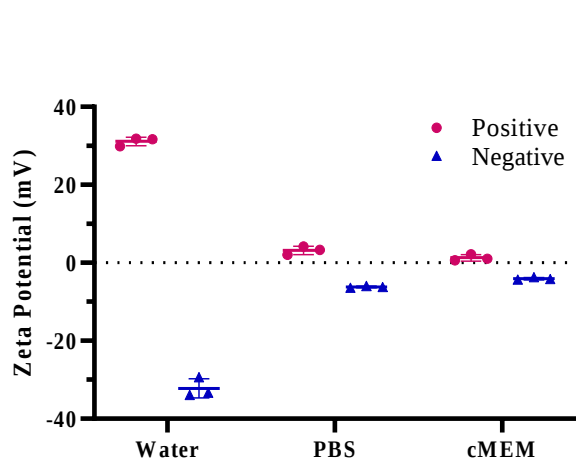


Why do positive nanoparticles usually have higher uptake than negative ones?

Feng Zhao



Preparing a corresponding liposome with the same amount of charges but negative

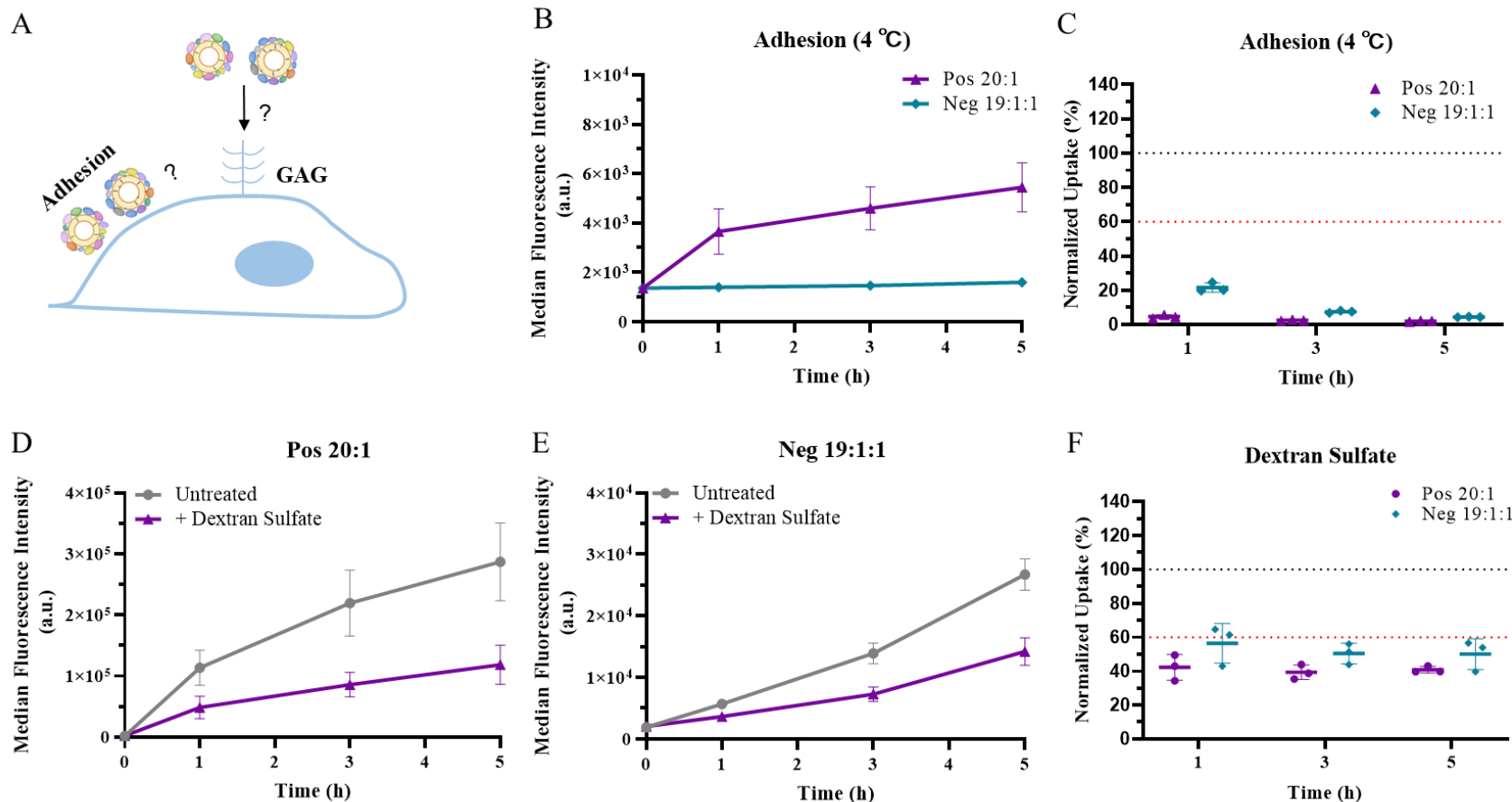


Why do positive nanoparticles usually have higher uptake than negative ones?

Feng Zhao



Common explanation: interaction with negatively charged proteoglycans on the cell membrane



→ Instead they both interact with heparan sulphate for uptake!

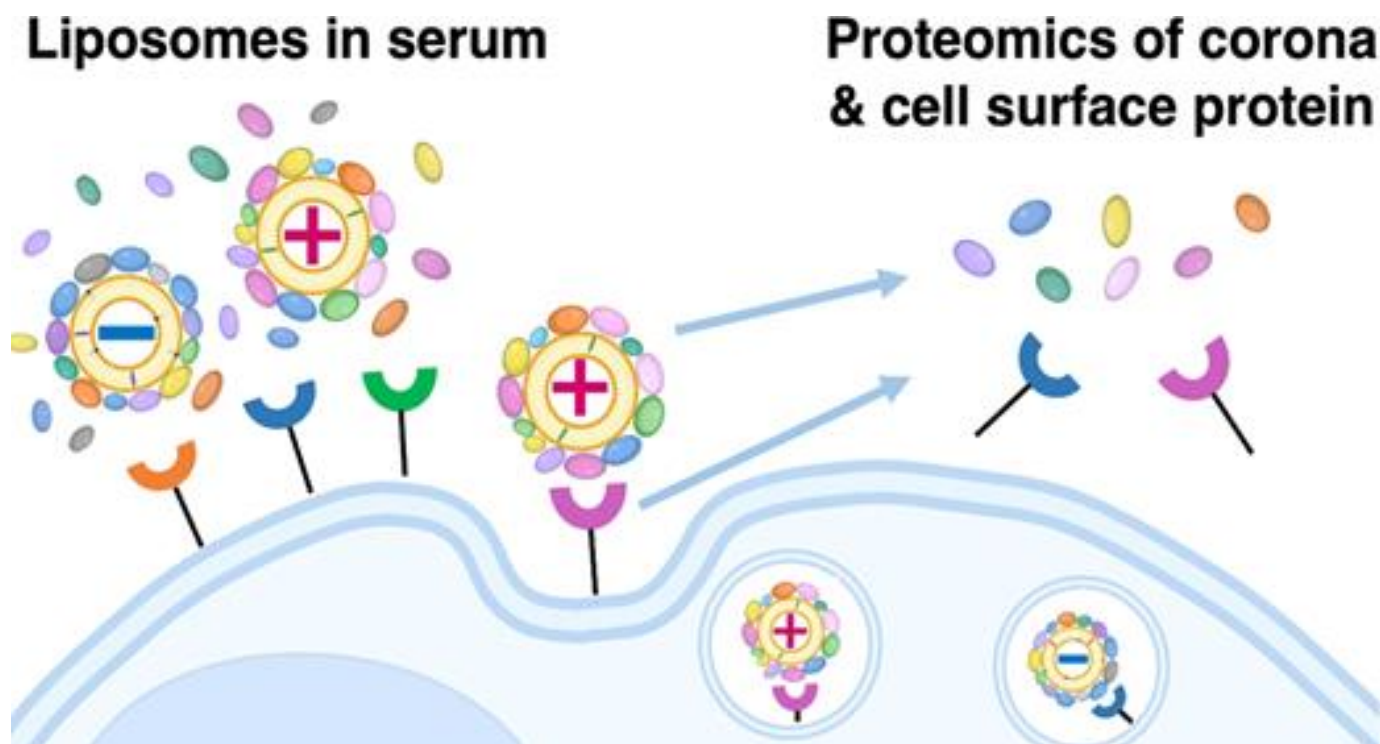
→ Slightly higher adhesion to cell membrane for positive liposomes

Comparison of Protein Coronas and Internalized Cell Surface Proteins of Positive and Negative Liposomes

Feng Zhao

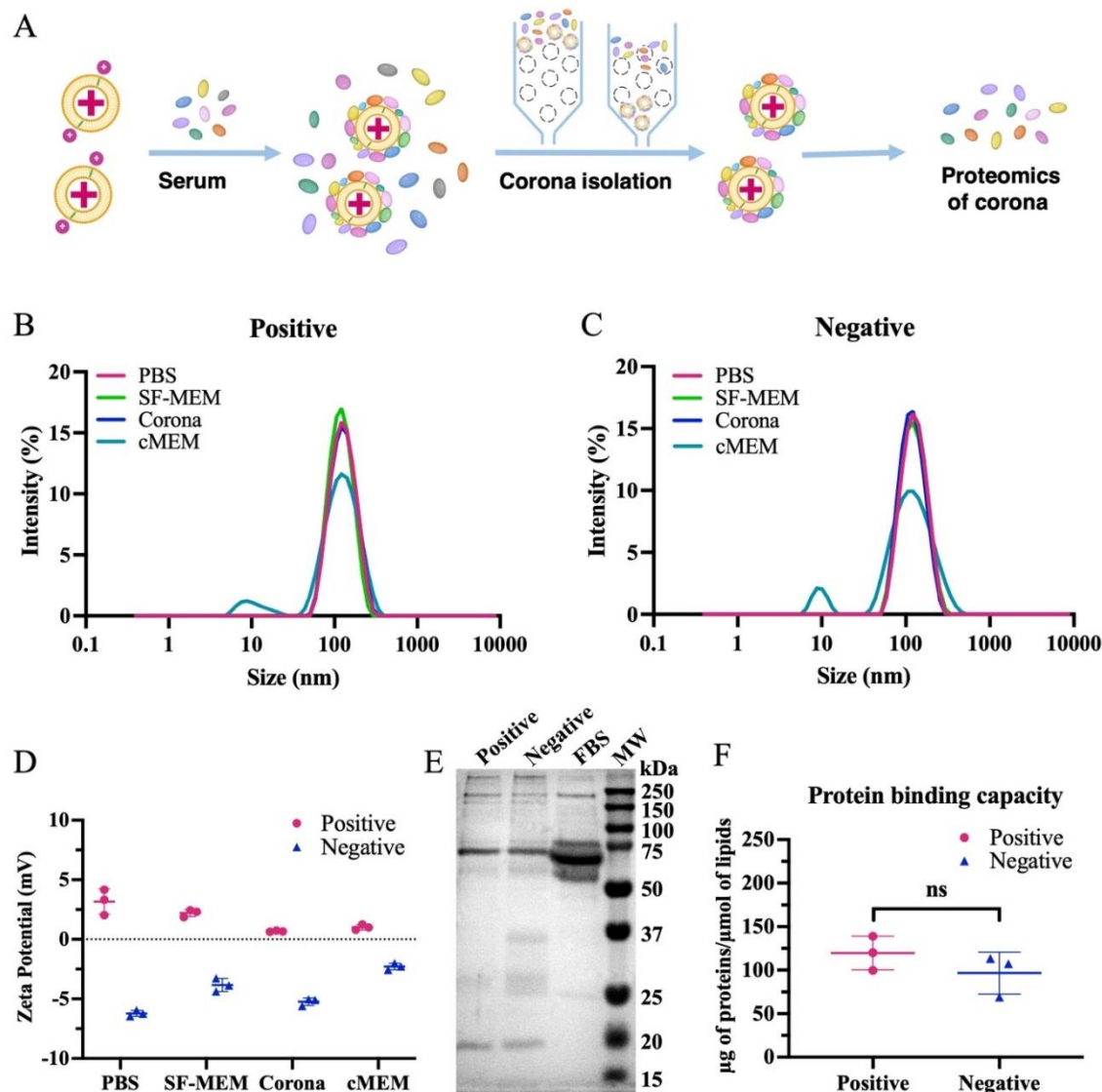


1. Protein corona adsorbed on the liposomes
2. Cell surface proteins involved in their internalization.



Comparison of Protein Coronas and Internalized Cell Surface Proteins of Positive and Negative Liposomes

Feng Zhao



Feng Zhao, Anna Salvati, et al. **ACS Nano**, 2026, **20** (27): 19225–19240

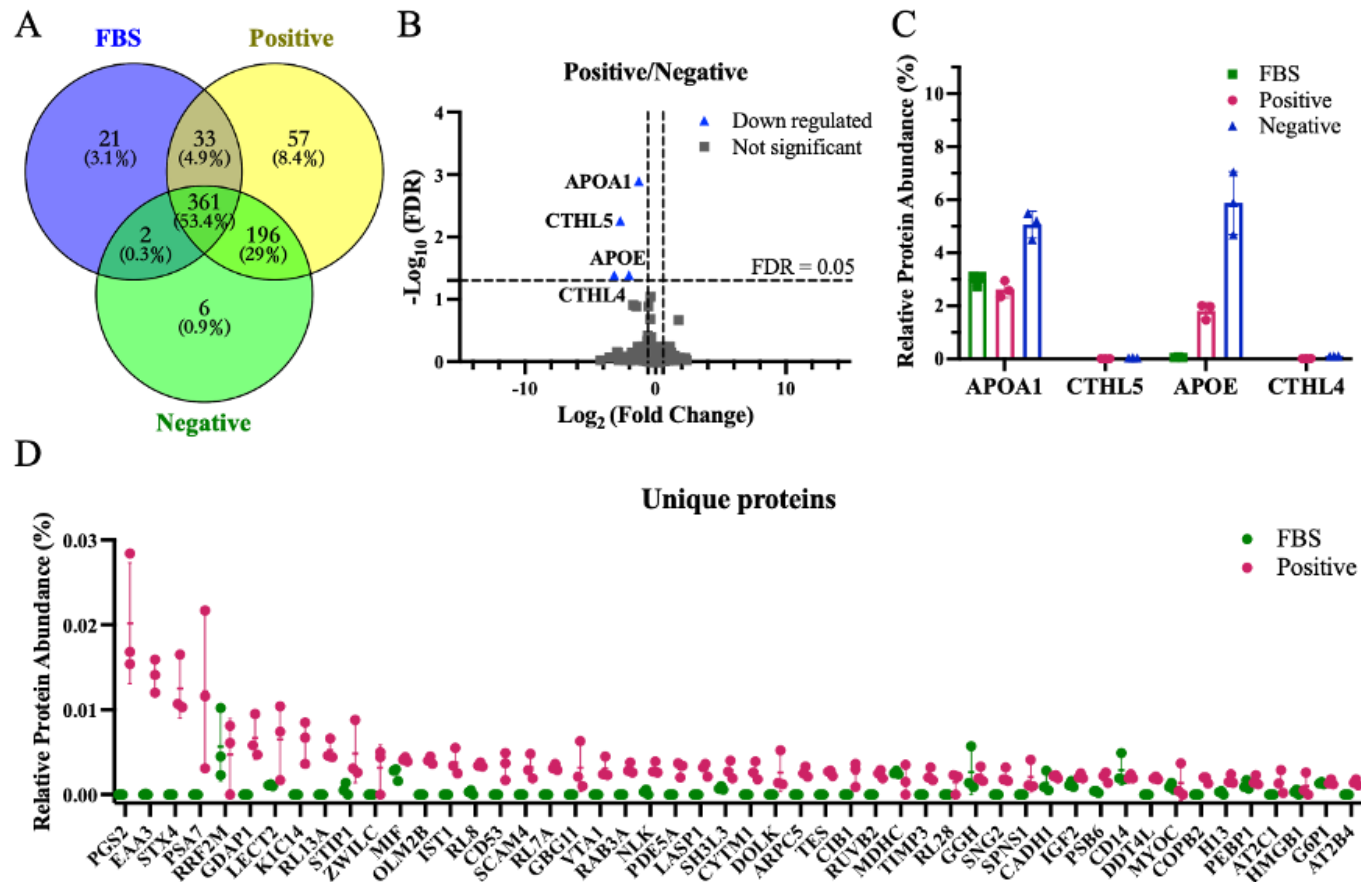
Comparison of Protein Coronas and Internalized Cell Surface Proteins of Positive and Negative Liposomes

Comparison of Protein Coronas and Internalized Cell Surface Proteins of Positive and Negative Liposomes

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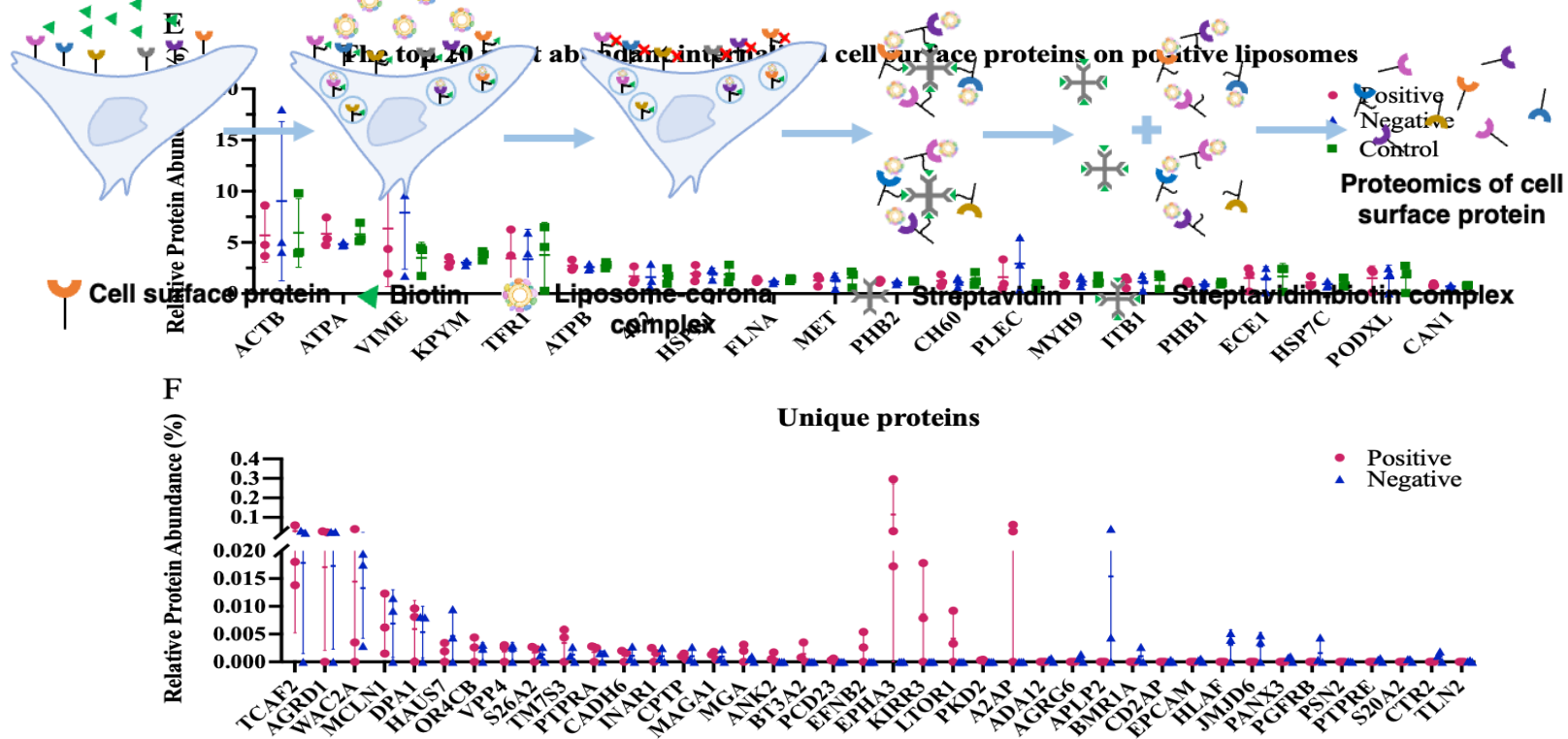


- Major differentially expressed proteins have low abundance
- Many of the most abundant proteins are common but differed in abundance
- **NB: not always few dominating corona proteins!**



Isolation of cell receptors using reversible cell surface biotinylation

Aldy Aliyandi, Anna Salvati, et al *Acta Biomaterialia*, 155, 507-520, 2022



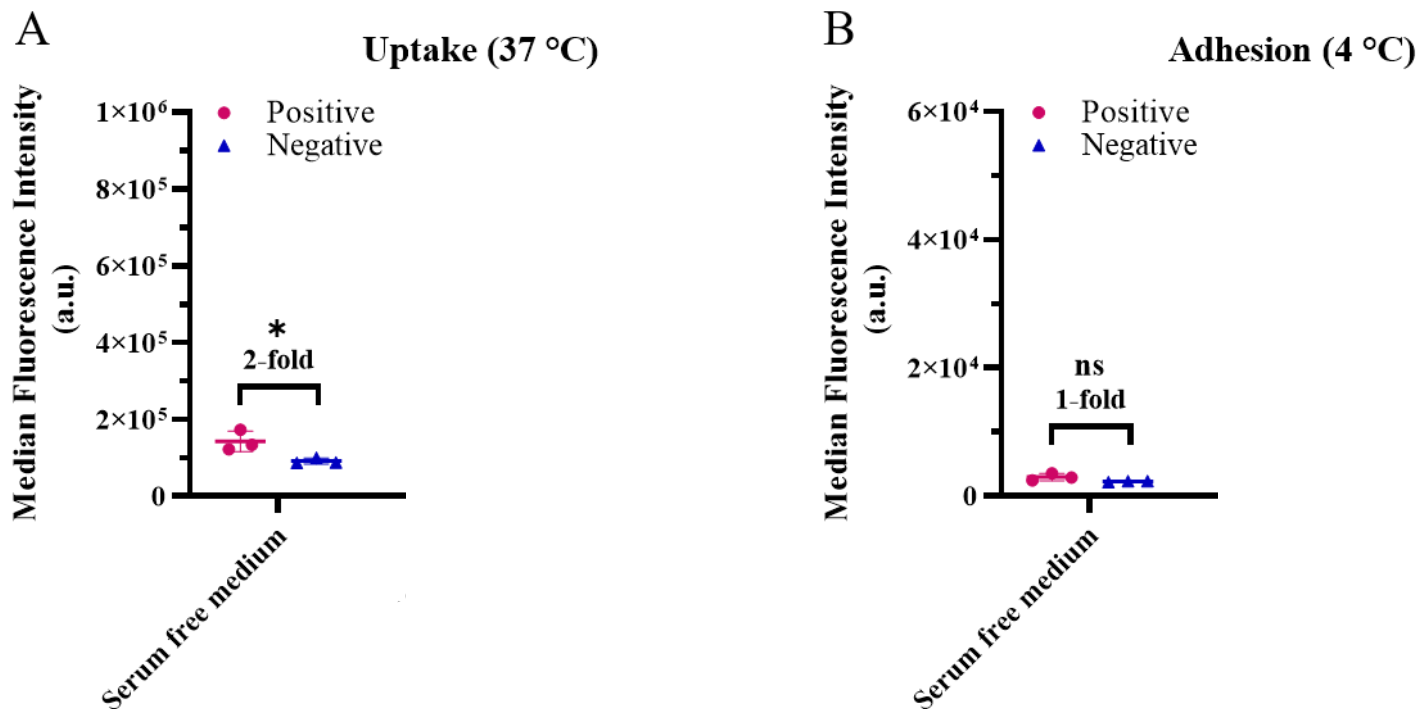
- Major differentially expressed proteins but low abundant.
- Most abundant proteins common in both systems with small differences
- **Not few dominating receptors! Much more complex pattern of subtle differences!**

Feng Zhao, Anna Salvati, et al. *ACS Nano*, 2026, **20** (27): 19225–19240

Comparison of Protein Coronas and Internalized Cell Surface Proteins of Positive and Negative Liposomes

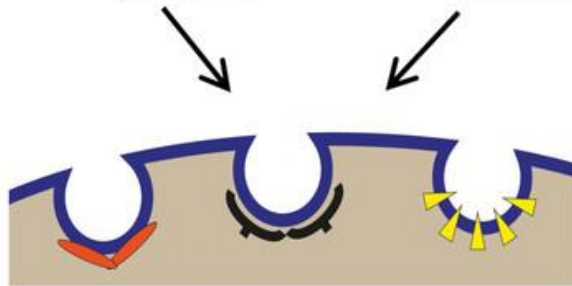
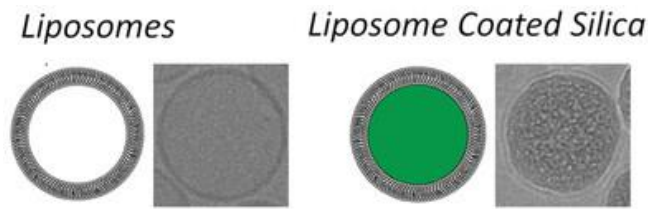


....does it matter?

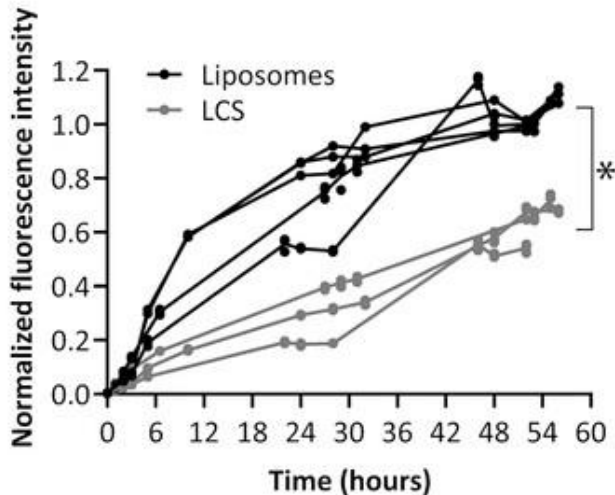


Electrostatic interactions alone cannot explain liposome uptake differences.
The uptake (and adhesion) of two liposomes is different only when proteins are present.

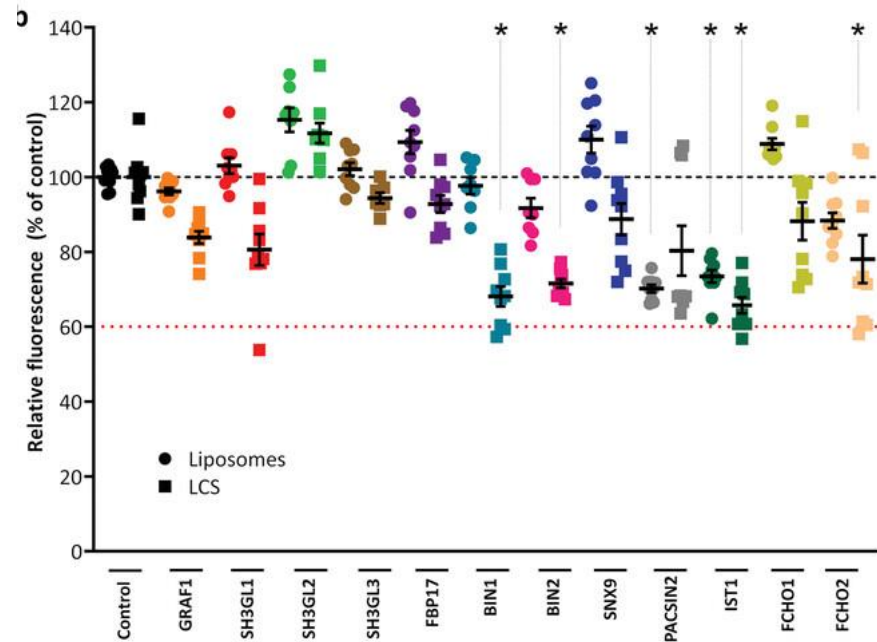
....another example: nanoparticle rigidity



Cell uptake & curvature-sensing proteins?



- Liposomes and liposome coated silica as a model
- Effect on uptake
- Involvement of specialized curvature sensing proteins (BAR domain proteins) in the internalization



... but does rigidity affect corona composition as well?

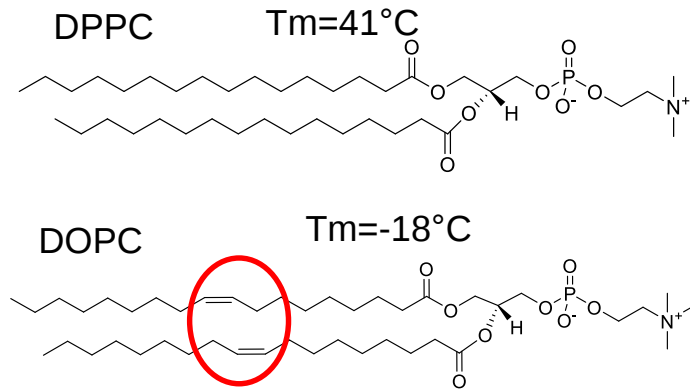
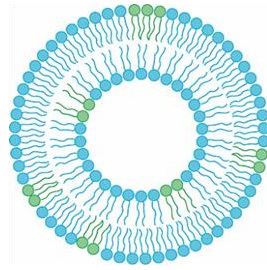
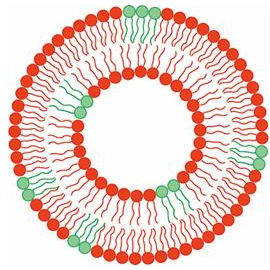
Changing lipids to tune liposome rigidity

Xinyu Ma

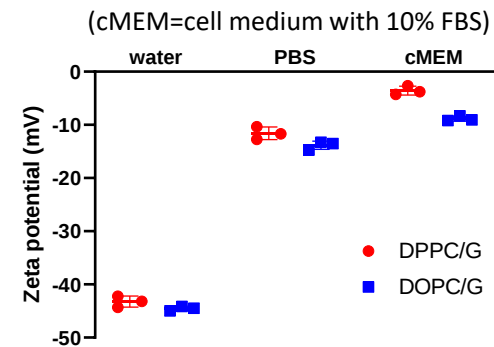
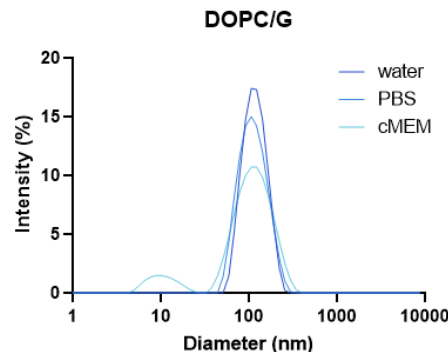
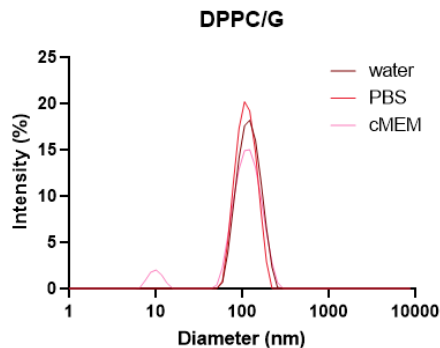


DPPC/G

DOPC/G



Adding as little as possible DOPG in both formulations to achieve stability



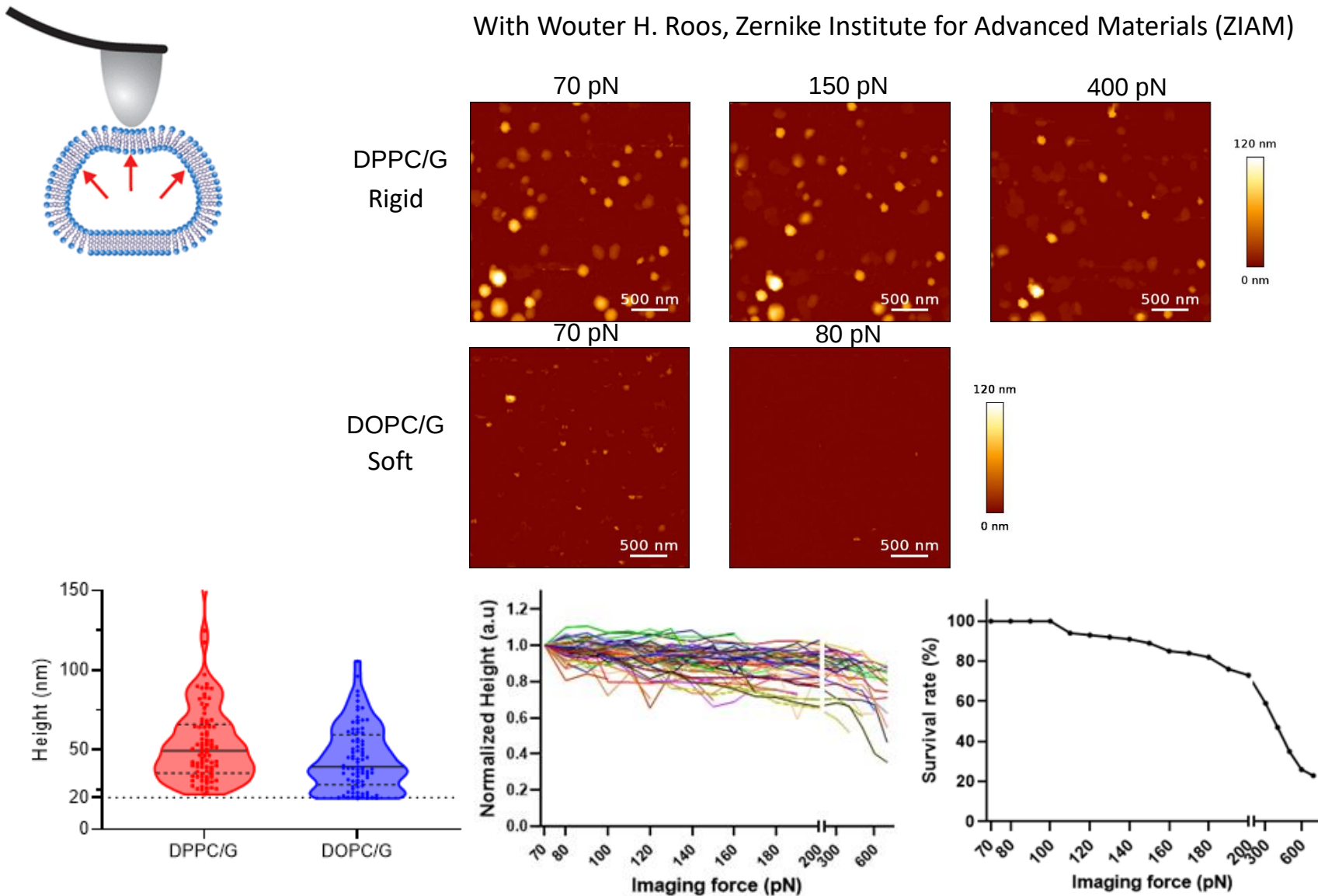
- Extensive optimization needed to obtain non-pegylated liposomes with solid bilayer that were stable in serum!
- Same size, same zeta potential, same surface composition, only acyl chain different!

AFM imaging at increasing imaging force

Xinyu Ma

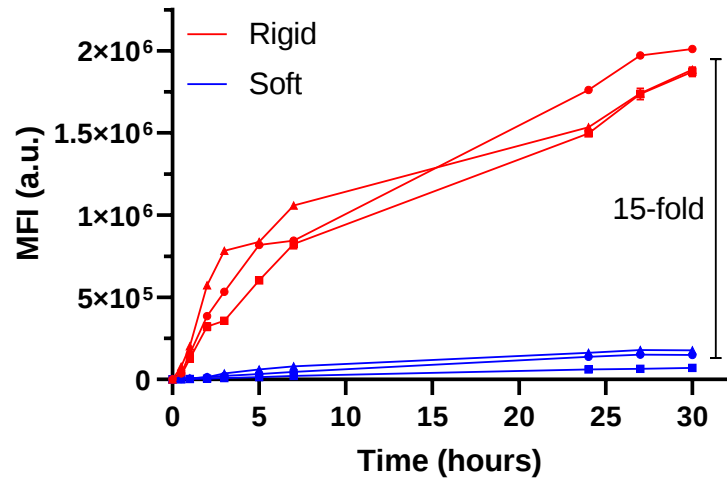


With Wouter H. Roos, Zernike Institute for Advanced Materials (ZIAM)

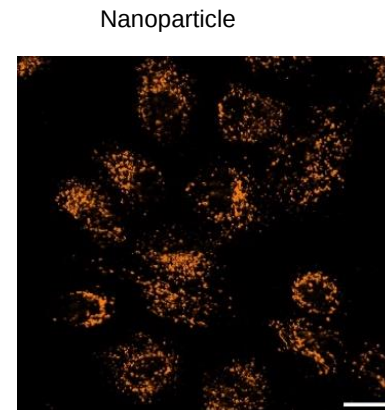


Rigid liposomes show higher uptake than soft liposomes

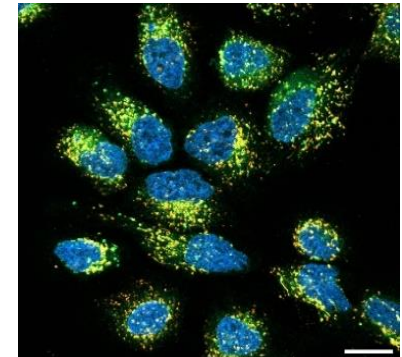
Xinyu Ma



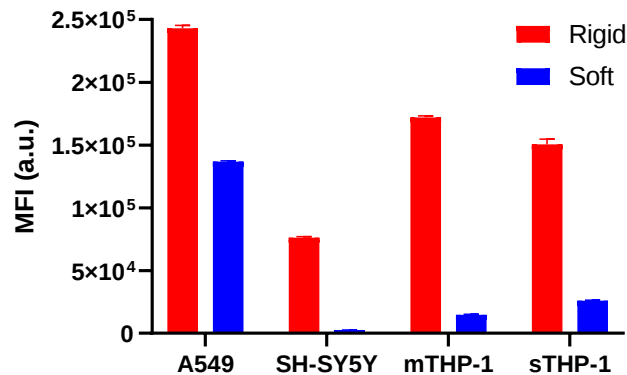
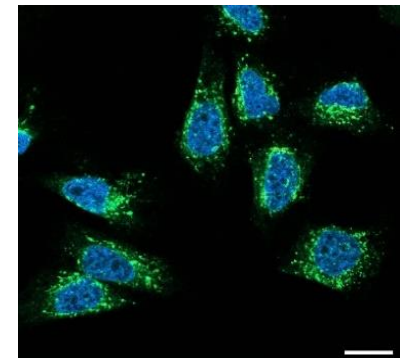
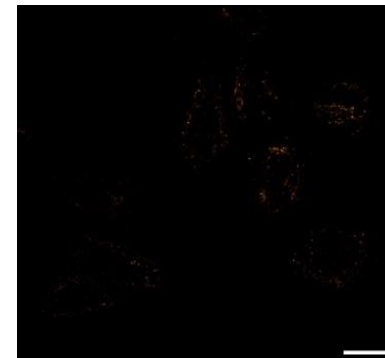
Rigid

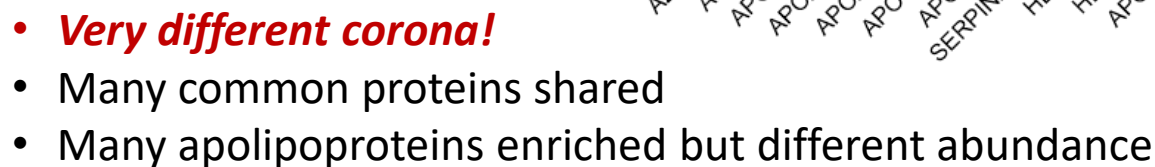


Merged



Soft





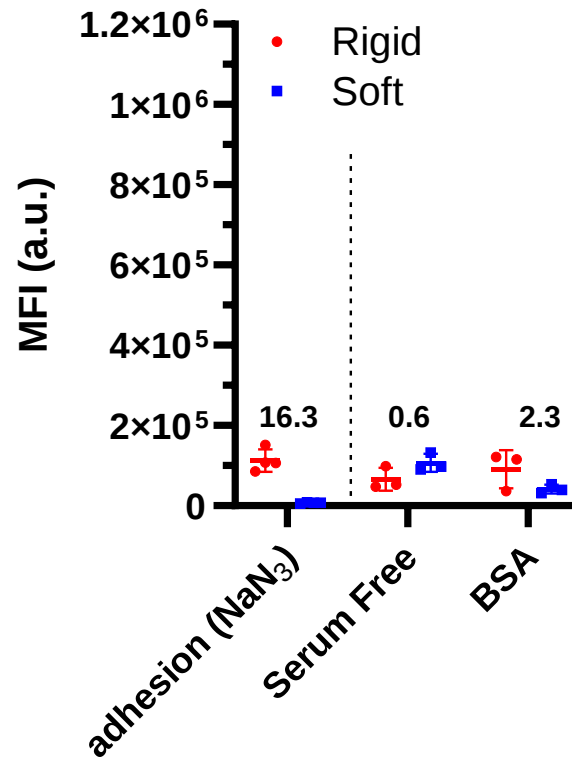
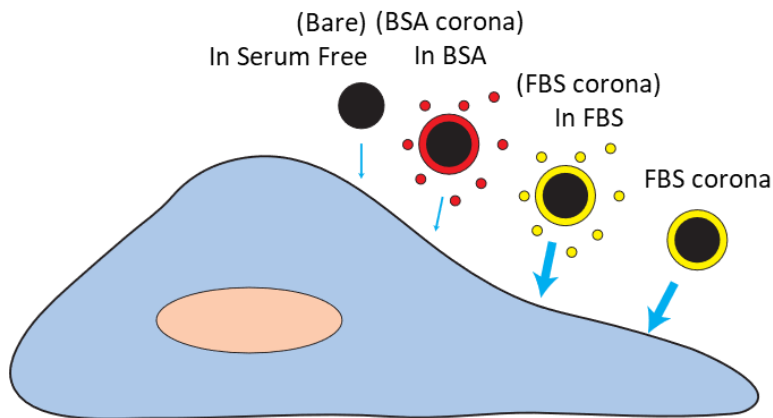
How much does the corona layer matter?

Xinyu Ma



Adding liposomes to cells in medium with different proteins

in different media

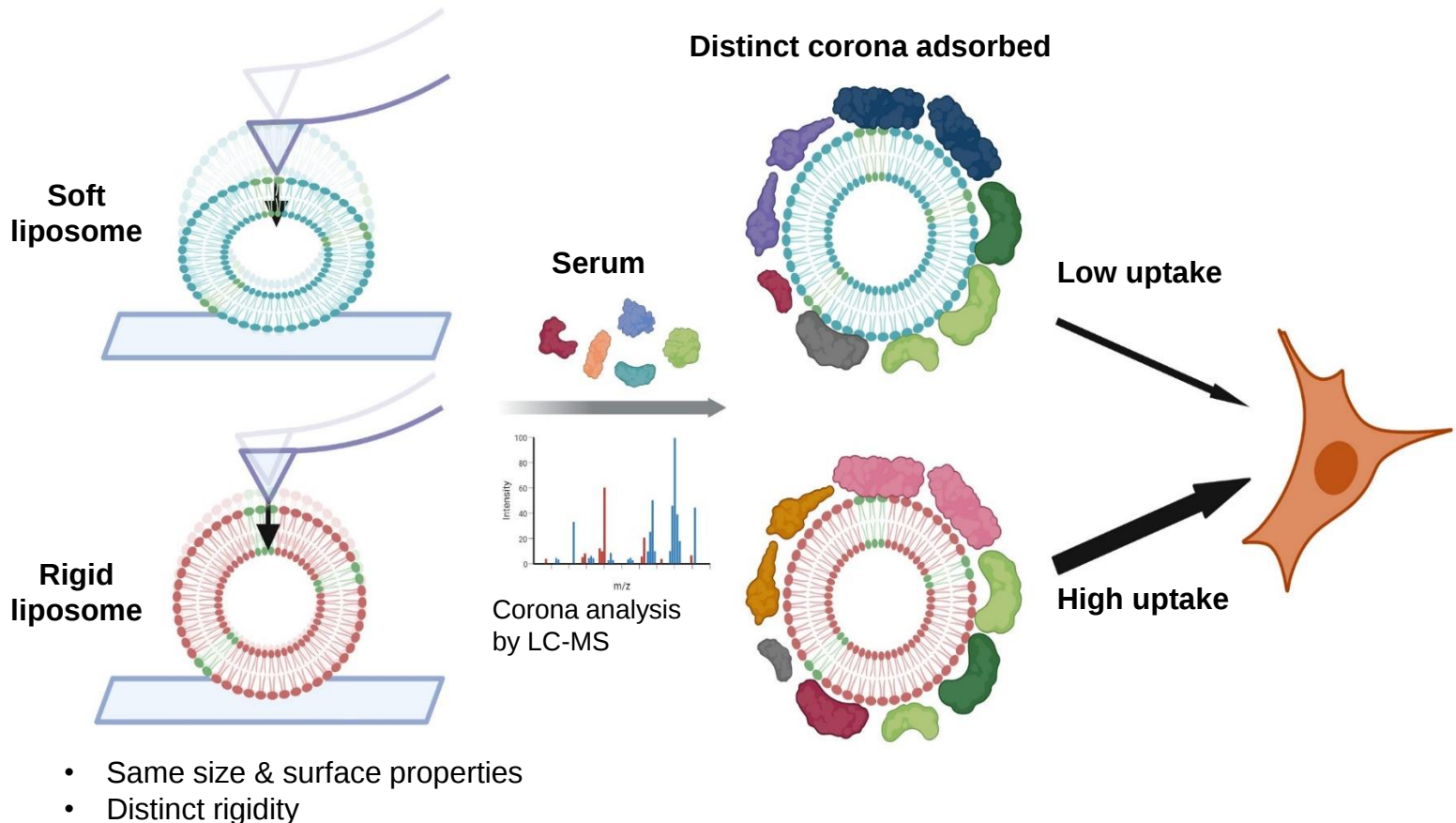


**Only when adding liposomes to cells
with serum, uptake is different!**

Effect of liposome rigidity on corona formation and its implications for uptake by cells

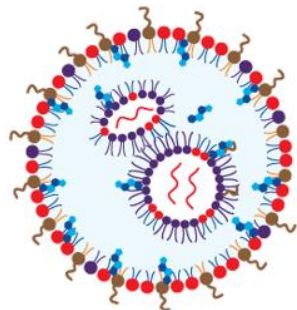
Xinyu Ma

Xinyu Ma, see poster #25

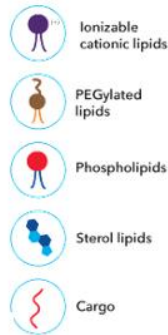


....another example: LNP and transfection efficiency

Heba Fayyaz

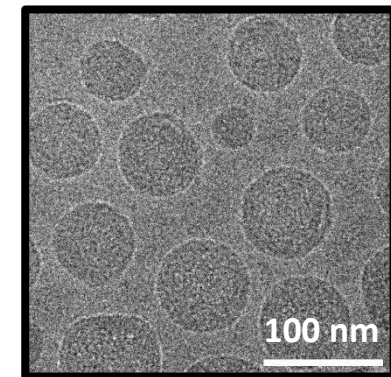
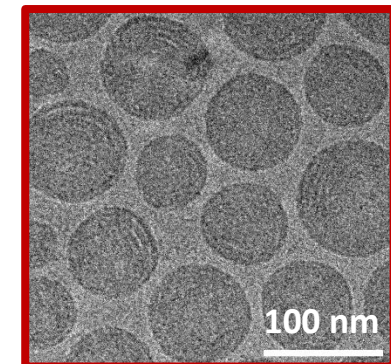
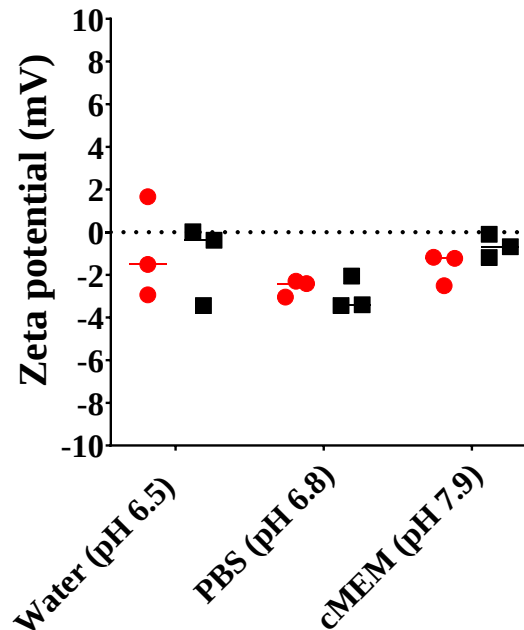
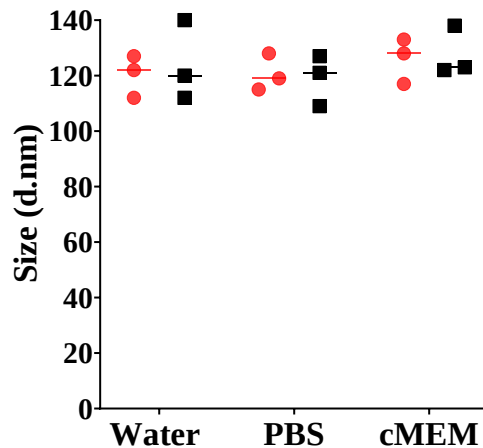


Lipid Nanoparticles



Molar ratio (Percent)				
DLin-MC3-DMA	DSPC	Cholesterol	DMG-PEG-2000	Dil
50	9.9	38.5	1.5	0.1

Poly-A loaded LNPs vs mRNA loaded LNPs



Heterogeneous cellular responses!

A second subpopulation upon incubation with LNPs!

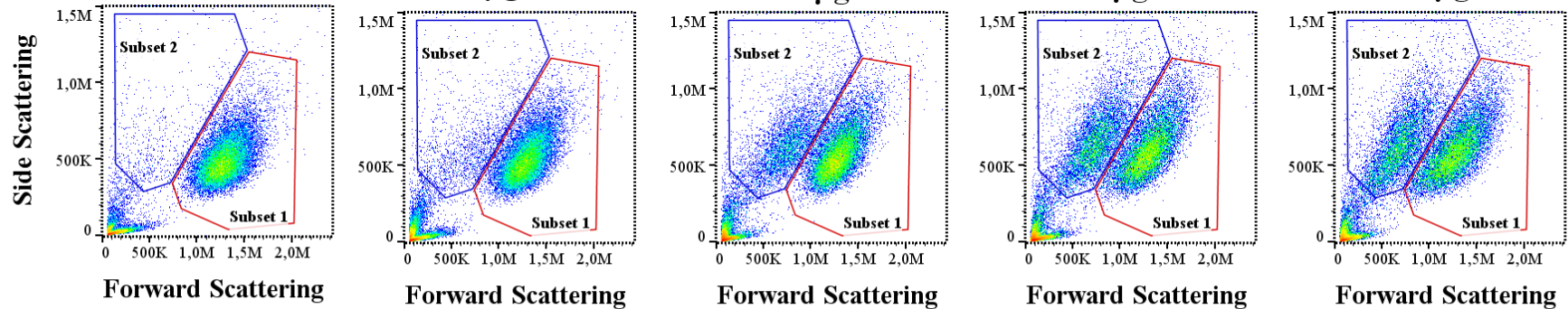
Untreated

3 $\mu\text{g/mL}$

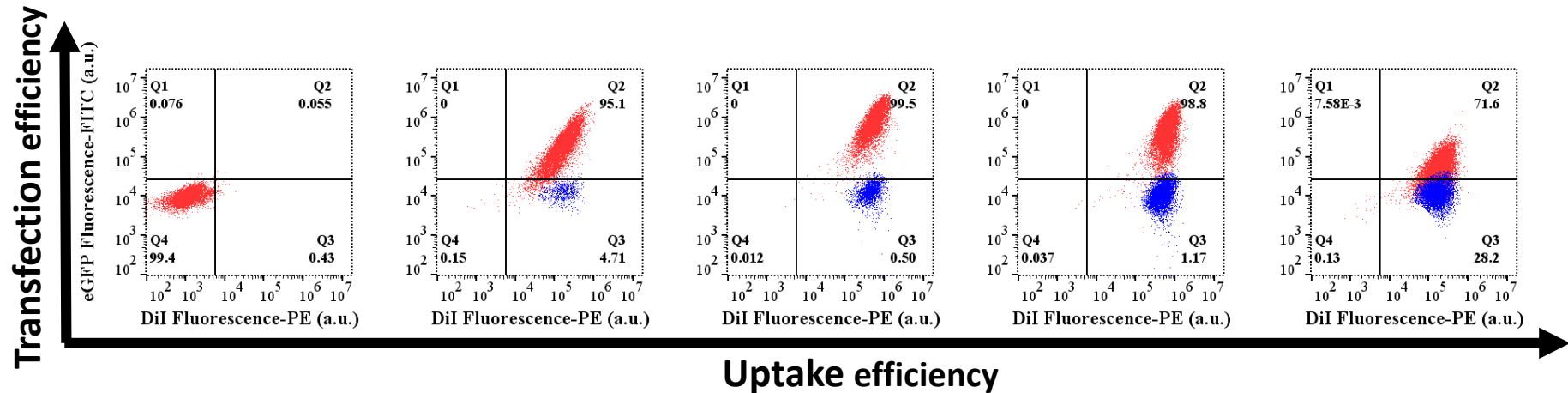
12.5 $\mu\text{g/mL}$

50 $\mu\text{g/mL}$

200 $\mu\text{g/mL}$

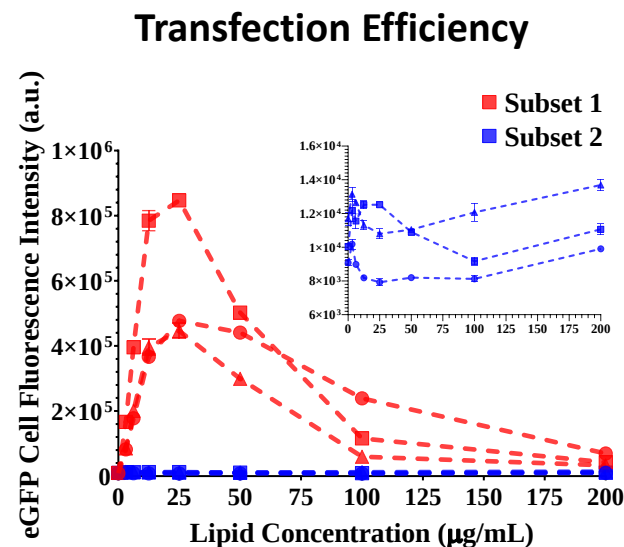
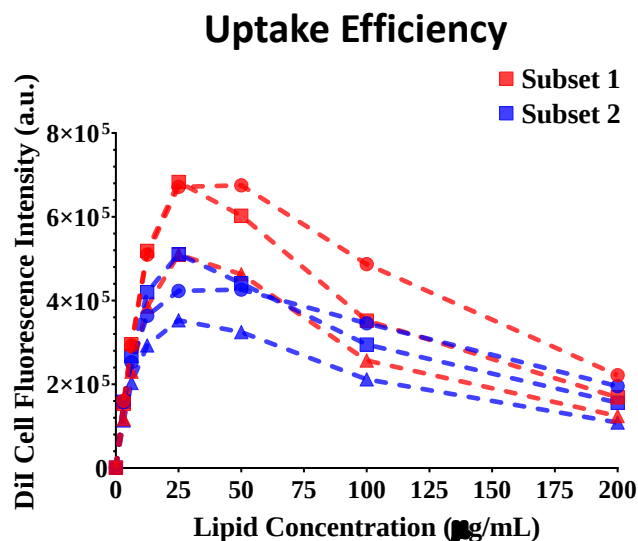


Subset 1 Subset 2



Non-monotonic dose response behaviour!

Heba Fayyaz



Endosomal Leakiness

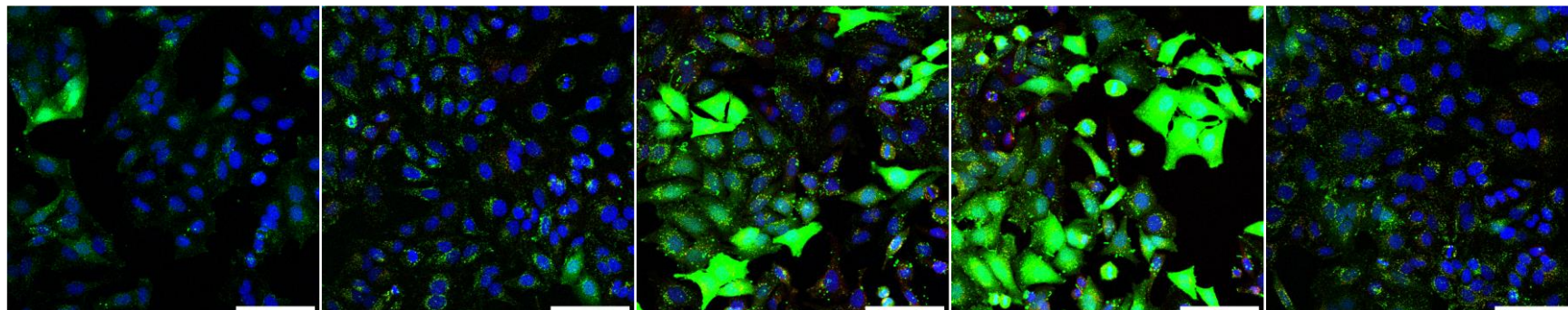
Untreated

2.5 $\mu\text{g/mL}$

10 $\mu\text{g/mL}$

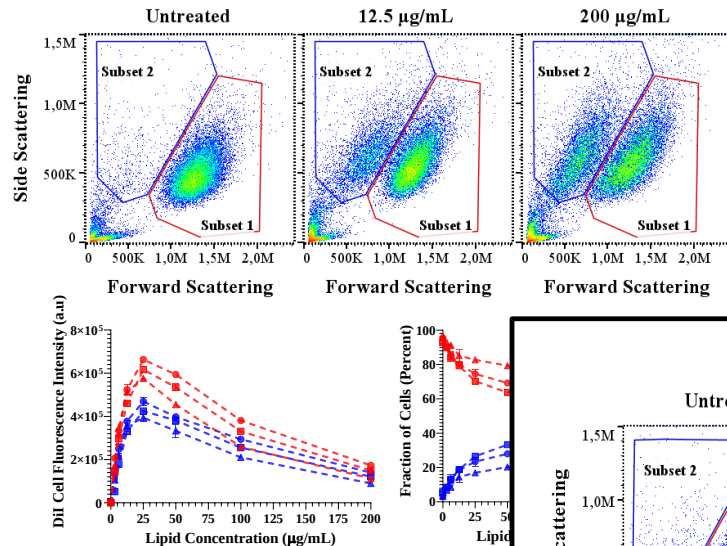
25 $\mu\text{g/mL}$

100 $\mu\text{g/mL}$

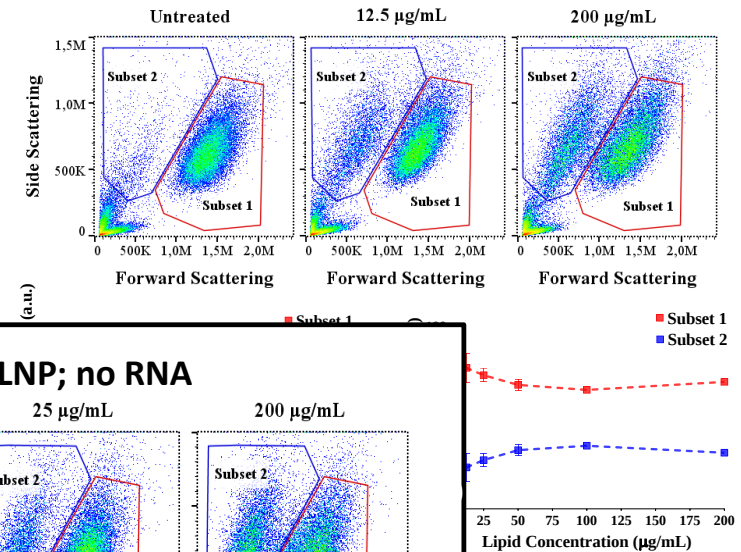




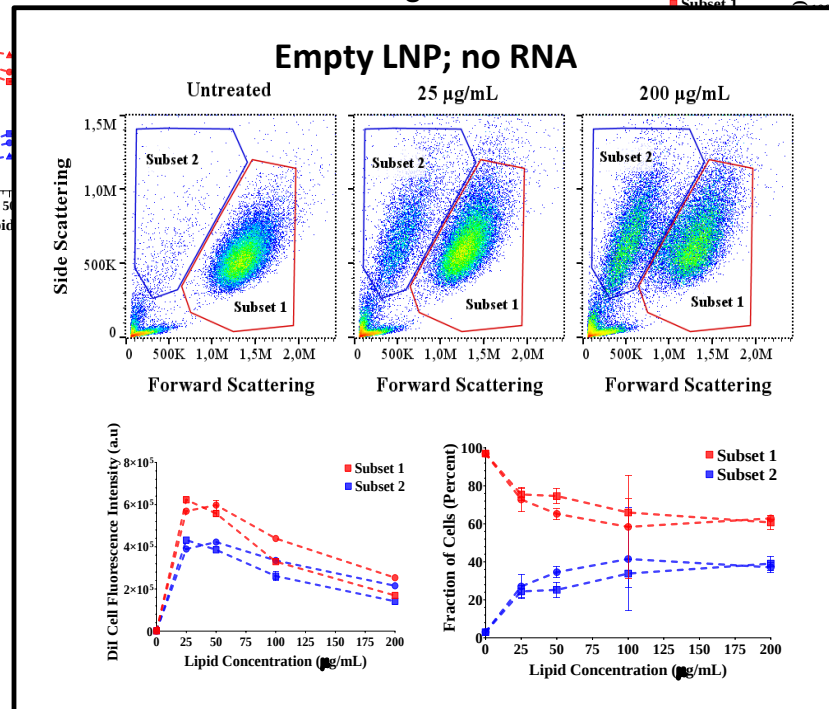
Poly-A LNPs at low RNA content



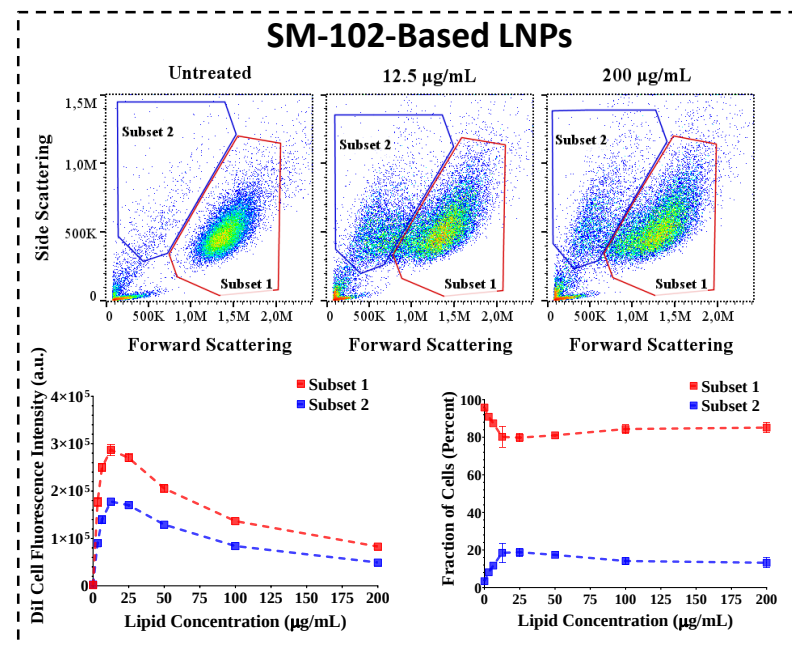
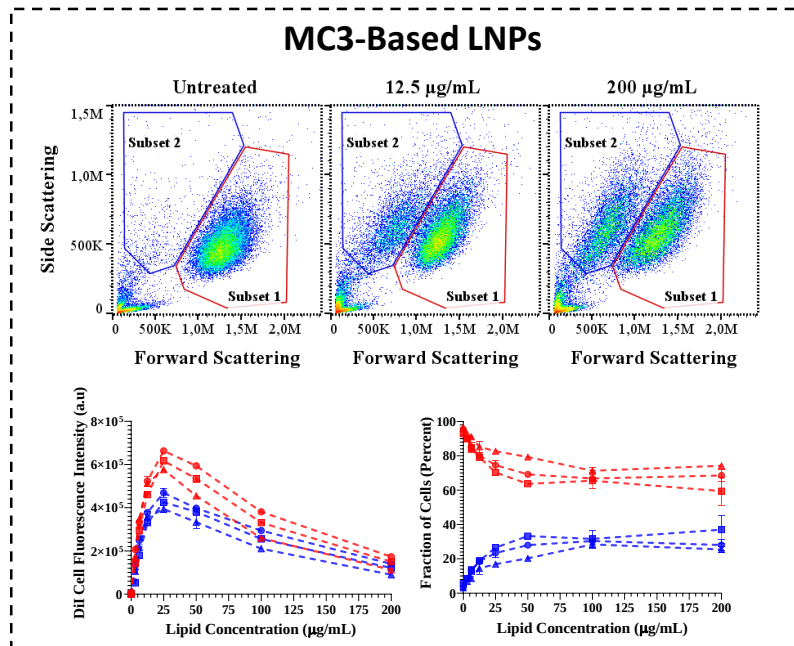
Poly-A LNPs at high RNA content



Empty LNP; no RNA



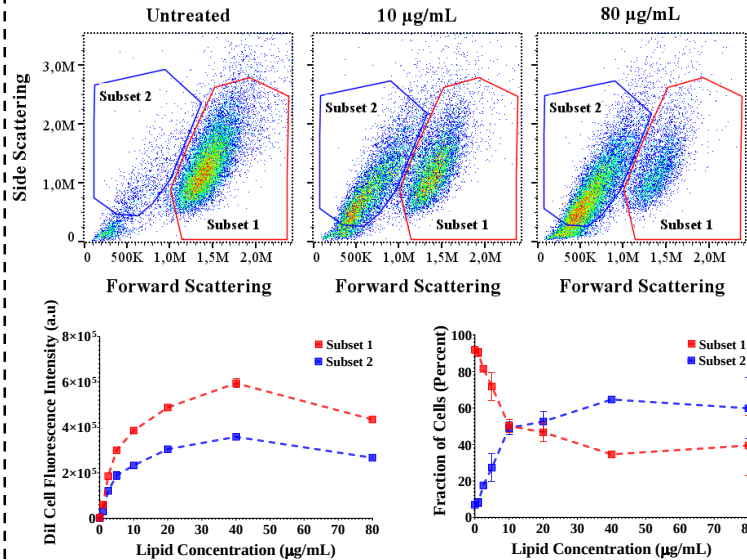
- Cargo independent
- Also for empty LNP!



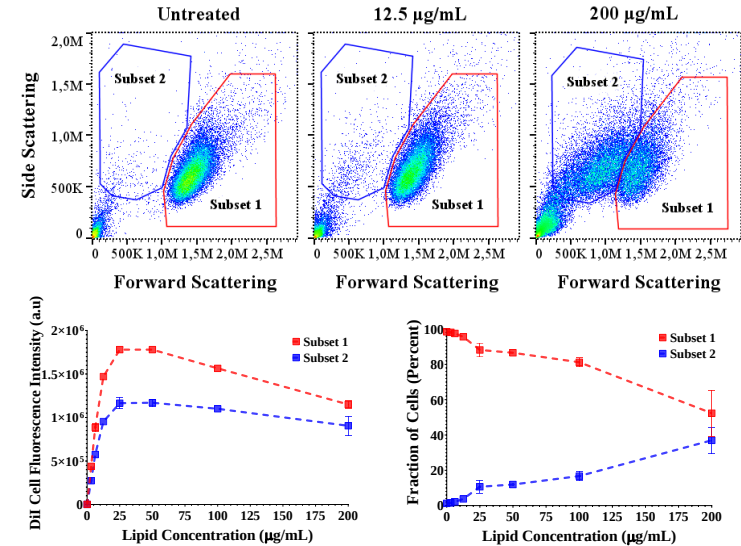
- Both with LNPs with MC3 and LNPs with SM-102



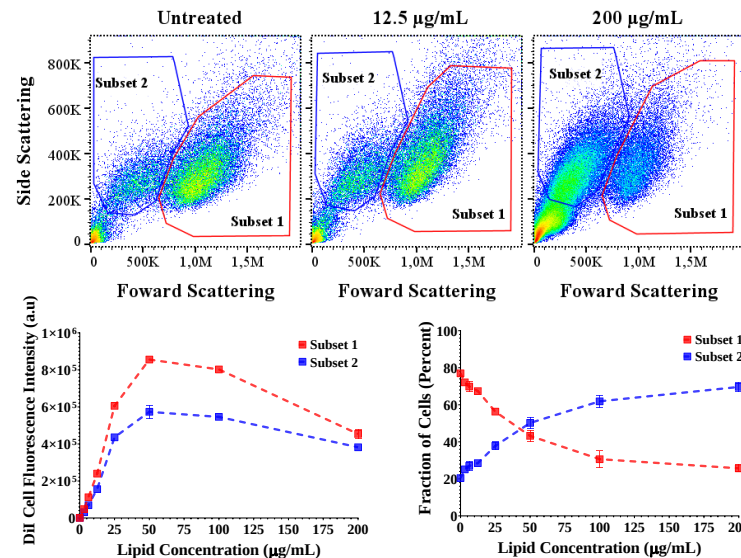
HuH-7



A549



HCT-15

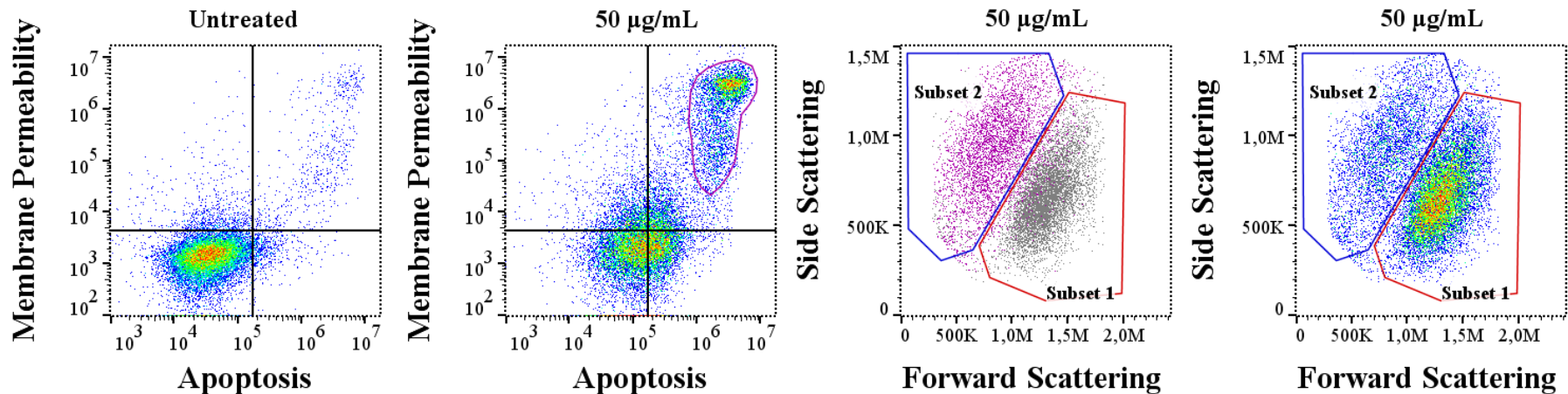


What is the second cell sub-population?

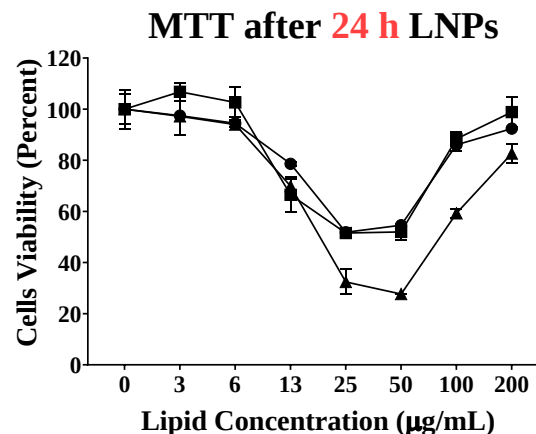
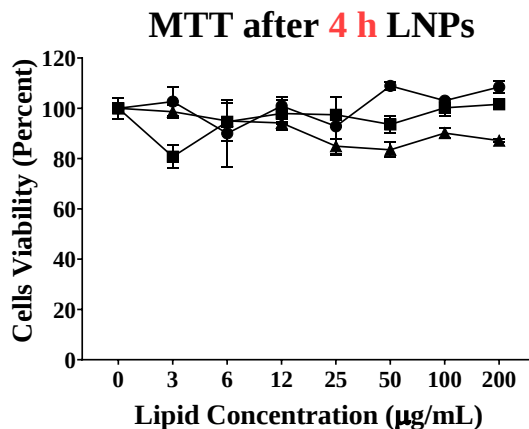
Heba Fayyaz



Subset 2 cells are PI- positive: a sub- population more sensitive to LNPs!



Cell viability correlated to uptake, not concentration

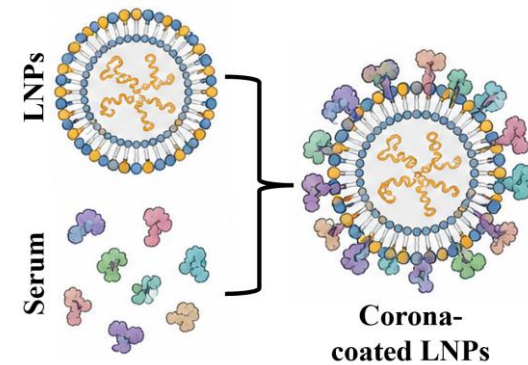


Why the non-monotonic uptake behavior?

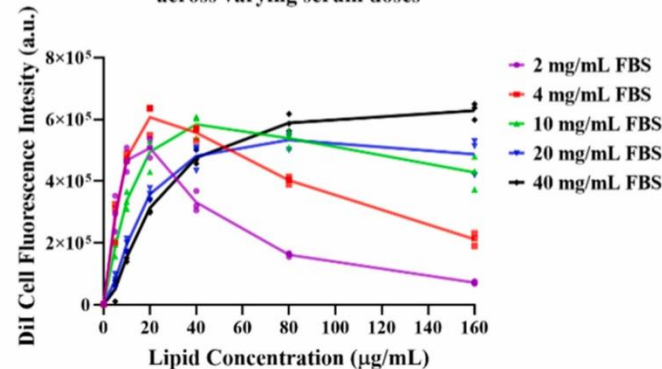
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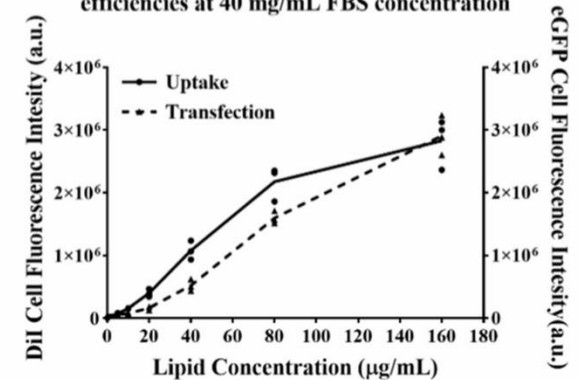
Adsorbed corona changes when changing LNP concentration



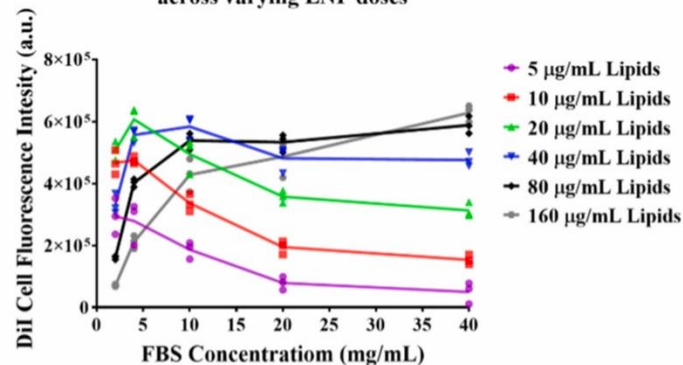
a Effect of LNP concentration on uptake efficiency across varying serum doses



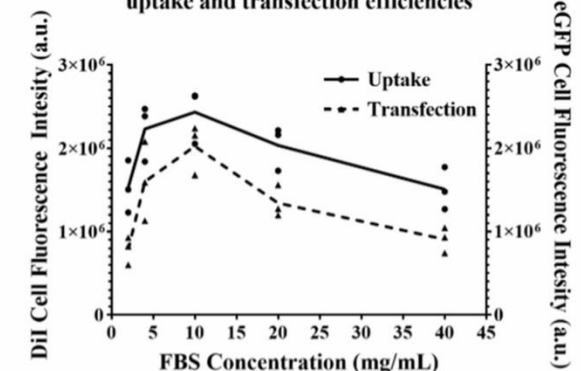
b Effect of LNP concentration on uptake and transfection efficiencies at 40 mg/mL FBS concentration



c Effect of serum concentration on LNP uptake efficiency across varying LNP doses



d Effect of serum concentration on LNPs of 80 μg/mL lipids uptake and transfection efficiencies



→ Uptake & transfection strongly modulated by exposure conditions (serum content)

Some conclusions



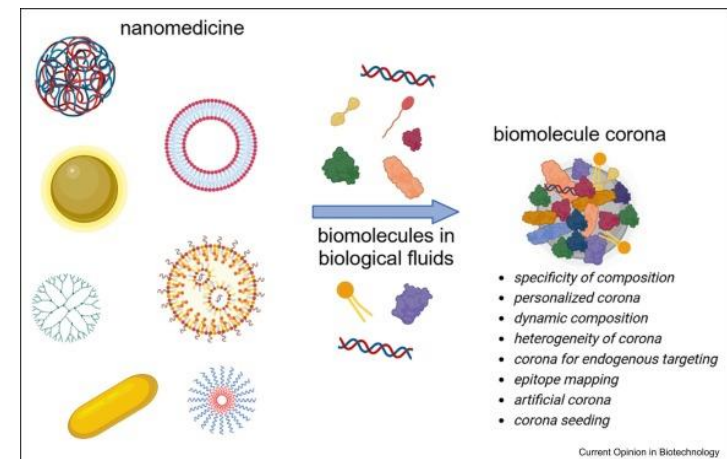
- Why do positive nanoparticles usually have higher uptake than negative ones?
- Effect of liposome rigidity on corona formation and its implications for uptake by cells
- Effect of protein corona on LNP uptake and transfection

→ *Nanomedicine interactions with biological fluids shape their cell behavior*

Salvati, *Current Opinion Biotech*, 2024, 87, 103101

The biomolecular corona of nanomedicines: effects on nanomedicine outcomes and emerging opportunities

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